The Role of Mast Cells in Non-Allergic Inflammation

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In 1878 Paul Ehrlich described a new type of granular cells that were mainly localized to the connective tissue compartment. These cells were prevalent in chronically inflamed human tissues. He named the cells "Mastzellen," meaning "well-fed cells" due to the high content of cytoplasmatic granules. These cells, better known today as mast cells, were initially considered to be part of the connective tissue. However, in the 1970s it was shown that their precursors were actually the hematopoietic stem cells. Mast cells circulate in the bloodstream and migrate into mucosal or connective tissues, where they undergo maturation into long-lived cells. These sites are the interface with the external environment (i.e., skin and mucosal surfaces), thus enabling MCs to respond rapidly to environmental stimuli, making them 'sentinels' of the immune system [1].

At first, MCs were studied in the context of allergic inflammation. For decades they were notable for their high affinity FC ϵ receptor-I, which after binding to immunoglobulin E forms the cell's antigen receptor. The interaction with antigen and the cross-linking of the FC ϵ RI causes degranulation of cytoplasmatic granules and the release of histamine, which increases vascular permeability and induces bronchoconstriction. The pathological consequences can be local, as in allergic rhinitis, or systemic as in anaphylaxis.

The discovery that MC granules contain preformed agents other than histamine (i.e., heparin, proteases, chondroitin sulfates and antimicrobial peptides) and their ability to selectively produce and release cytokines, chemokines, growth factors and lipid mediators (i.e., prostaglandins and leukotrienes) has led many investigators to believe that MCs participate in various biological responses other than IgE-mediated allergic inflammation. This review will focus on some of the recent advances in understanding these cells heterogeneity, mainly their role in non-allergic inflammation [2].

Mast cells and innate immunity

Both the innate and the adaptive arms of the immune system participate in host defense against infections. The unique localization of MCs to the host-environment interface, being a common site of pathogen invasion, and their ability to react to

MC = mast cells FC&R = FC& receptor Ig = immunoglobulin a large variety of physical, biological and chemical stimuli, raised the idea that these cells might play a role in innate immunity against infections [3,4].

MCs were initially associated with host defense against parasites since these pathogens cause IgE-associated responses. Studies have shown that intestinal parasitic infections were associated with MC hyperplasia and subsequently the release of proteases that results in expulsion of parasites by disrupting the intestinal epithelial barrier [1,5-7]. Moreover, studies of MC-deficient or IgE-deficient mice showed that these factors had a role in protection against intestinal worms [1,4,6,8]. The role of MCs in the defense against intracellular parasites was also shown in several infection models, including: malaria, Toxoplasma gondii, Trypanosoma brucei, Giardia lamblia and Leishmania [5,9-11].

In 1996, two groups published reports that have changed our understanding of MC biology [12,13]. They studied the ability of MC-deficient mice (lacking the MC growth factor receptor-c-kit) to fight Klebsiella peritonitis. Both groups found that normal mice overcame the bacterial infection, whereas MC-deficient mice died as a result of it. This was attributed to the rapid MC secretion of tumor necrosis factor-alpha following the inoculation of the bacteria, resulting in an augmented neutrophil response. Their findings were the first to clearly demonstrate that MCs have an important role in innate immunity, a role that is not related to parasitic infections. Shortly after, other researchers demonstrated MCs' roles in host defense against additional bacteria such as Escherichia coli and Mycoplasma pneumoniae [14-17].

MCs can be activated by host-derived signals or directly by the pathogen. The former include activated products of the complement system and endogenous peptides, which are formed quickly and in large quantities in response to infection [1,6]. For example, it has recently been shown that MCs were activated by Fv protein, an endogenous protein released by the liver during viral hepatitis [2,4,18]. Interestingly, the interaction was mediated through the Fc receptor. An example of the latter is the interaction between CD48 on the MCs and the fimbrial antigen FimH expressed by several gram-negative bacteria [2,15,17,19]. It has also been shown that MCs were activated following interaction between dengue virus and the FcγRII, resulting in the release of specific mediators that were different from those released after interaction with bacterial products [20-22].

The toll-like receptor family comprises single membranespanning receptors that recognize conserved molecules expressed by different pathogens but not by the host. These molecules include factors such as bacterial peptidoglycan, lipopolysacharide, dsRNA and bacterial DNA. TLRs are considered among the key players that alert the immune system to the presence of pathogens. Since they are expressed on MCs, it was reasonable to believe that MCs participate in the recognition phase of innate immune responses [23,24]. For example, studies have shown that viruses activate MCs through interaction with TLR3 (that recognize dsRNA), resulting in the secretion of interferon-alpha, which inhibits viral replication and recruits other immune cells such as natural killer cells and macrophages [24,25].

Mast cells and adaptive immunity

There is a growing body of evidence that bidirectional interactions between MCs and T lymphocytes have a major role in adaptive immunity [26]. MCs are able to phagocyte bacteria, process its antigens, and present it to T lymphocytes in the context of major histocompatibility complex class I and II, thus serving as antigen-presenting cells [5,27,28]. During induction of an immune response. MCs migrate into lymph nodes where they further secrete chemokines and cytokines that induce lymph node hypertrophy and aggregation of additional lymphocytes [26,29]. $TNF\alpha$ induces T lymphocyte recruitment to the lymph node while interleukin-6 promotes these cells' activation [4,30]. On the one hand, most of the cytokines secreted by MCs induce Th2 differentiation, thereby escalating the allergic immune response. On the other hand MCs can secrete IL-12 and INFy that support Th1 response, suggesting in fact that MCs are able to regulate the equilibrium between Th1 and Th2 responses [5].

MCs regulate T lymphocytes' specific immune responses indirectly by modulation of dendritic cells [5,31,32]. They promote recruitment, maturation and migration of immature dendritic cells from the circulation to the lymph nodes, where they present antigens to T lymphocytes [23,29,33]. Moreover, MCs have a regulatory effect on B cells through expression of MHC class II, stored in exosomes that release through exocytosis [5,34]. MCs express a wide variety of surface receptors and adhesion molecules that can be implicated in the co-stimulation process during the adaptive immune responses and enable them to interact with other inflammatory cells. Examples of such receptors are intercellular adhesion molecule-1, β 2-integrins and CD40 ligand [2,34-36].

Snake bites and bee stings

For many years MCs were believed to contribute to the complications caused by snake bites and bee stings through the release of tissue-damaging molecules. These molecules promote an increase in vascular permeability, local inflammation, disturbance of the clotting and the fibrinolysis systems, and eventually might lead to shock and death. In 2004, Maurer and colleagues [37] published a study where they investigated the association between MCs

TLR = toll-like receptor

 $TNF\alpha$ = tumor necrosis factor-alpha

IL = interleukin

INFy = interferon-gamma

MHC = major histocompatibility complex

and endothelin-1 [37]. ET-1 is an endogenous vasoconstrictor peptide that participates in the vascular changes occurring during sepsis. It is also known to activate MCs by binding to the ET-1 receptors, leading to degranulation and release of mediators. Interestingly, the authors showed that MCs promote the degradation of ET-1, thereby increasing survival during acute bacterial peritonitis [37].

ET-1 has high homology (> 70%) to sarafotoxin, the most potent toxic components of the venom of the Israeli mole viper (Seraph Ein Gedi). This raised the question whether MCs have a protective role in envenomation. In 2006, Metz and Galli [38] challenged this question. They found that MC-deficient mice were more susceptible to hypothermia and death as a result of injection of either sarafotoxin or complete Israeli mole viper venom. compared to normal mice or MC-deficient mice engrafted with normal MCs. Moreover, they showed that the levels of sarafotoxins in the peritoneal cavity of these mice were significantly lower than in the control group, suggesting that MCs have a role in reducing the toxin levels. They generated two groups of mice that contained MCs that either expressed or lacked the ET-1 receptor and found that those lacking the receptor failed to clear sarafotoxins and died quickly, indicating that sarafotoxins activate MCs, at least in part, through binding to this receptor.

In their search for a potential mechanism by which MCs reduced the venom toxicity, they found that carboxypeptidase A, a protease found in MC granules, was responsible for degrading the venom. Normal mice given an inhibitor of CPA, or MC-deficient mice engrafted with MCs expressing an inhibitory RNA that silenced CPA, died within an hour after injection of sarafotoxins or whole venom. Further studies showed that MCs had a protective role against other venoms that did not contain ET-like peptides such as the American pit vipers, the western diamondback rattlesnake, and the southern copperhead [38]. It was also reported that MCs provided protection against honeybee venom, although it is still unclear if CPA is the major anti-venom agent in this case [39]. Despite the fact that human MCs are different from mice MCs, it is reasonable to assume that they share similar protective roles [40].

Conclusions

It was not too long ago that the role of MCs was restricted to allergic inflammation. However, in the last two decades MCs have emerged as one of the most important factors at both the innate and the adaptive arms of the immune system. There is no doubt that we are only at the beginning of the journey towards better understanding of MC biology and extensive research is definitely needed.

References

- Metz M, Maurer M. Mast cells key effector cells in immune responses. Trends Immunol 2007;28:234–41.
- Bachelet I, Levi-Schaffer F, Mekori YA. Mast cells: not only in allergy. Immunol Allergy Clin North Am 2006;26:407–25.

ET-1 = endothelin-1

CPA = carboxypeptidase A

- Mekori YA, Metcalfe DD. Mast cells in innate immunity. Immunol Rev 2000:173:131–40.
- 4. Dawicki W, Marshall JS. New and emerging roles for mast cells in host defence. Curr Opin Immunol 2007;19:31–8.
- Stelekati E, Orinska Z, Bulfone-Paus S. Mast cells in allergy: innate instructors of adaptive responses. J Imbio 2007;212:505–19.
- Marshall JS. Mast cell responses to pathogens. Nat Rev Immunol 2004;4:787–99.
- Dermott JR, Bartram RE, Knight PA, Miller HR, Garrod DR, Grencis RK. Mast cells disrupt epithelial barrier function during enteric nematode infection. Proc Natl Acad Sci USA 2003;100: 7761–6.
- Onah DN, Nawa Y. Mucosal mast cell derived chondroitin sulphate levels in and worm expulsion from FcRγ-knockout mice following oral challenge with Strongyloides venezuelensis. J Vet Sci 2004:5:221–6.
- Henderson WR, Chi EY. The importance of leukotrienes in mast cell mediated Toxoplasma gondii cytotoxicity. J Infect Dis 1997;177: 1437–43
- Ben-Rashed M, Ingram GA, Pentreath VW. Mast cells, histamine and the pathogenesis of intestinal damage in experimental Trupanosoma brucei infections. Ann Trop Med Parasitol 2003;97:803–9.
- Birdi M, Vouldoukis I, Mossalayi MD, et al. Evidence for direct interaction between mast cells and Leishmania parasites. Parasite Immunol 1997;19:475–83.
- Echtenacher B, Mannel DN, Hultner L. Critical protective role of mast cells in a model of acute septic peritonitis. Nature 1996; 381:75–7.
- Malaviva R, Ikeda T, Ross E, Abraham SN. Mast cell modulation of neutrophil influx and bacterial clearance at sites of infection through TNF-alpha. Nature 1996;381:77–80.
- Kulka M, Fukuishi N, Rottem M, Mekori YA, Metcalfe DD. Mast cells, which interact with Escherichia coli, up-regulate genes associated with innate immunity and become less responsive to FcεRImediated activation. J Leukoc Biol 2006;79:339–50.
- Thankavel K, Madison B, Ikeda T, et al. Localization of a domain in the FimH adhesin of Escherichia coli type 1 fimbriae capable of receptor recognition and use of a domain-specific antibody to confer protection against experimental urinary tract infection. J Clin Invest 1997;100:1123–36.
- Xu X, Zhang D, Lyubynska N, et al. Mast cells protect mice from Mycoplasma pneumonia. Am I Respir Crit Care Med 2007;173:219–25.
- Malaviya R, Ikeda T, Abraham SN. Contribution of mast cells to bacterial clearance and their proliferation during experimental cystitis induced by type 1 fimbriated E. coli. Immunol Lett 2004; 91:103–11
- Genovese A, Borgia G, Bouvet JP, et al. Protein Fv produced during viral hepatitis is an endogenous immunoglobulin superantigen activating human heart mast cells. Int Arch Allergy Immunol 2003;132:336–45.
- Abraham S, Shin J, Malaviya R. Type 1 fimbriated Escherichia colimast cell interactions in cystitis. J Infect Dis 2001;183(S1):51–5.
- King CA, Marshall JS, Alshurafa H, Anderson R. Release of vasoactive cytokines by antibody-enhanced dengue virus infection of a human mast cell/basophil line. J Virol 2000;74:7146–50.
- King CA, Anderson R, Marshall JS. Dengue virus selectively induces human mast cell chemokine production. J Virol 2002; 76:8408–19
- Brown MA, King CA, Marshall JS, Anderson RP. A dominant role for Fcγ.RII in antibody-enhanced dengue virus infection of human mast cells and associated CCL5 release. J Leukoc Biol 2006;80: 1242–50.

- Heib V, Becker M, Warger T, et al. Mast cells are crucial for early inflammation, migration of Langerhans cells and CTL responses following topical application of TLR7 ligand in mice. Blood 2007; 110:946–53.
- Kulka M, Alexopoulou L, Flavell RA, Metcalfe DD. Activation of mast cells by double-stranded RNA: evidence for activation through Toll-like receptor 3. J Allergy Clin Immunol 2004;114: 174–82.
- Orinska Z, Bulanova E, Budagian V, Metz M, Maurer M, Bulfone-Paus S. TLR3-induced activation of mast cells modulates CD8+ T-cell recruitment. Blood 2005;106:978–87.
- Sayed BA, Brown MA. Mast cells as modulators of T-cell responses. Immunol Rev 2007;217:53–64.
- 27. Malaviya R, Twesten N, Ross E, Abraham SN. Mast cells process bacterial antigens through a phagocytic route for class I MHC presentation to T cells. I Immunol 1996:156:1490–6.
- 28. Warbrick EV, Taylor AM, Botchkarev VA, Coleman JW. Rat connective tissue-type mast cells express MHC class II: up-regulation by IFN-gamma. Cell Immunol 1995;163:222–8.
- Jawdat DM, Rowden G, Marshall JS. Mast cells have a pivotal role in TNF-independent lymph node hypertrophy and the mobilization of langerhans cells in response to bacterial peptidoglycan. J Immunol 2006;177:1755–62.
- Wang HW, Tedla N, Lloyd AR, Wakefield D, Neil PH. Mast cell activation and migration to lymph nodes during induction of an immune response in mice. J Clin Invest 1998;102:1617–26.
- Metz M, Grimbaldeston MA, Nakae S, Piliponsky AM, Tsai M, Galli SJ. Mast cells in the promotion and limitation of chronic inflammation. *Immunol Rev* 2007;217:304–28.
- 32. Galli SJ, Nakae S, Tsai M. Mast cells in the development of adaptive immune responses. *Nat Immunol* 2005;6:135–42.
- Demeure CE, Brahimi K, Hacini F, et al. Anopheles mosquito bites activate cutaneous mast cells leading to a local inflammatory response and lymph node hyperplasia. J Immunol 2005;174: 3932–40.
- Galli SJ, Kalesnikoff J, Grimbaldeston MA, Piliponsky AM, Williams CM, Tsai M. Mast cells as "tunable" effector and immunoregulatory cells: recent advances. Annu Rev Immunol 2005;23: 749–86.
- 35. Weber S, Babina M, Feller G, Henz BM. Human leukaemic mast cells (HMCS-1) and normal skin mast cells express beta 2-integrins: characterization of beta 2-integrins and ICAM-1 on HMCS-1 cells. Scand | Immunol 1997;45:471–81.
- Stahl JL, Cook EB, Graziano FM, Barney NP. Human conjunctival mast cells: expression of Fc epsilonRI, c-kit, ICAM-1, and IgE. Arch Ophthalmol 1999;117:493–7.
- Maurer M, Wedemeyer J, Metz M, et al. Mast cells promote homeostasis by limiting endothelin-1-induced toxicity. Nature 2004; 432:512–16.
- 38. Metz M, Piliponsky AM, Chen CC, et al. Mast cells enhance resistance to snake and honeybee venoms. *Science* 2006;313:526–30.
- Rivera J. Snake bites and bee stings: the mast cell strikes back. Nature Med 2006;12:999–1000.
- Bischoff SC. Role of mast cells in allergic and non-allergic immune responses: comparison of human and murine data. Nature Rev Allergy Immunol 2007;7:93–104.

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